

FAT: a novel domain in PIK-related kinases

Phosphatidylinositol kinases are found in all eukaryotes and serve important functions in phosphatidylinositol (PI) signaling pathways¹. In addition to the PI-kinase domain, most of these proteins have a number of accessory domains, usually involved in protein-protein interactions, that specify the role in a given pathway. A few examples of such domain organizations are shown in Fig. 1a. Recently, a new subfamily of the PI-kinase superfamily has emerged, called PIK-related^{2,3}. Although these proteins are large (2000–4000 amino acids), they only share similarity to classical PI

kinases in the ~300-amino-acid kinase domain.

Members of the PIK-related family appear functionally distinct, as none of them has been shown to phosphorylate lipids, such as PI; instead, many have Ser/Thr protein kinase activity^{4–7}. Despite this functional disparity, we will refer to this domain as the PI-kinase domain. Many PIK-related proteins are involved in cell-cycle checkpoint control [e.g. ATM, ATR, DNA-PK, ESR1 and Rad3 (reviewed in Ref. 8)]. Dysfunction can result in a range of diseases, including immunodeficiency, neurological disorder and cancer⁹.

It has previously been noted that members of the PIK-related family share a unique motif at the extreme C terminus¹⁰, which we call FATC. However, it has proved difficult to define shared domains in the large N-terminal portions. Although

sequence similarity between various members extends upstream of the PI-kinase domain, it usually tapers off in an irregular way. Several different N-terminal domain configurations have been hinted at in schematic diagrams, but no domain was supported by a multiple alignment^{5,11–14}. Tentative assignments of a leucine zipper and a DNA-polymerase-processivity-factor-binding site (P-site) have been made in the N-terminal region¹⁵. Although these regions were shown to be functionally important, the putative leucine-zipper motif is unlikely to be a true leucine zipper, given the atypical composition of the non-leucine residues¹⁶. The P-site motif is uncertain too, as five out of nine amino acids are never observed at the same position in other known cases¹⁷.

A group of proteins distantly related to PI kinases comprises the TRRAP

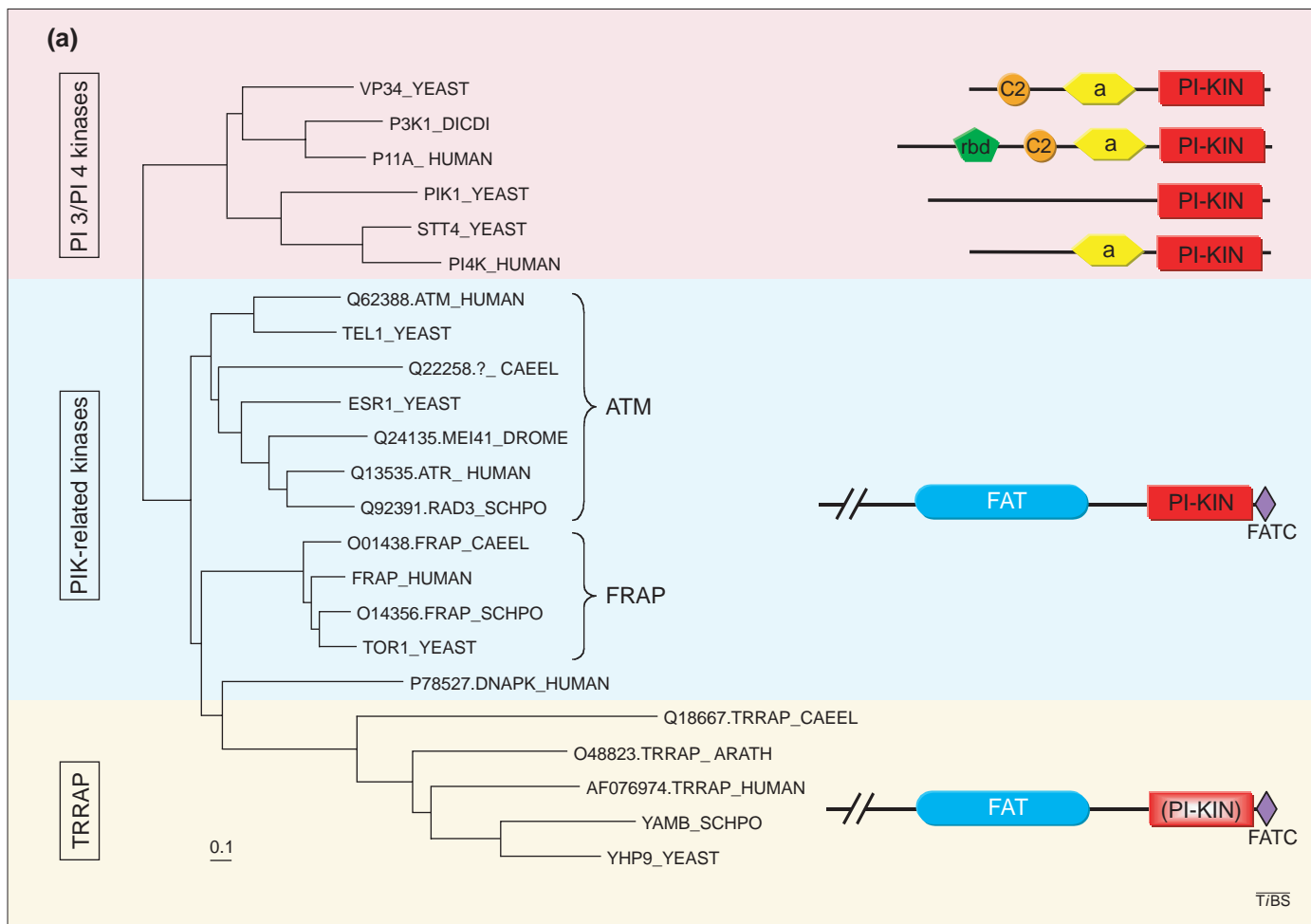


Figure 1

(a) Modular architecture and tree based on the PI-kinase domain (PI-KIN in red rectangle) of several representatives of the PI 3-/PI 4-kinases and all known PIK-related and TRRAP proteins. Within the PIK-related and TRRAP subfamilies, all members share the same domain architecture, as indicated by the tree sections colored blue and yellow. The FAT domain is only present in the FRAP, ATM and TRRAP subfamilies and always coexists with the FATC domain. Other domains in the PI 3-/PI 4-kinases are named according to Pfam (Ref. 21) but are shown without the 'PI3K' prefix. These are PI3K_C2 (PF00792), C2-like domain in PI kinases; PI3Ka (PF00613), accessory (PIK) domain; PI3K_rbd (PF00794), ras-binding domain. For more information on these domains, see <http://www.cgr.ki.se/Pfam>. The tree was generated by PHYLO_WIN (Ref. 22) using the neighbor-joining method with pairwise gap removal and PAM distances, from a multiple alignment of the PI-kinase domain generated by CLUSTALW (Ref. 23). The PI-kinase domain in TRRAP is denoted (PI-KIN) and drawn with a hollow because it lacks the catalytic residues despite sequence homology. (b) Multiple alignment of the FAT domain in all known sequences. Residues were colored by Belvu based on average pairwise BLOSUM62 score exceeding 0.3 (gray shade) or 1.3 (blue). Sequence names are either the Swissprot ID or a TREMBL accession number followed by a dot and a name mimicking a Swissprot ID, except for TRRAP_HUMAN for which only an EMBL accession number is available. Because Q22258 could not be classified unambiguously we used a question mark.

(b)

Table of protein sequence motifs with columns for accession number, organism, motif sequence, and position. Includes entries for Q62388.ATM HUMAN, Q22258.? CAEEL, and Q24135.ME141 DROME.

proteins¹⁸. Their PI-kinase domain lacks the catalytic residues and indeed, none of them has been shown to possess kinase activity. The TRRAP proteins contain regions of similarity to PIK-related proteins in the neighboring regions of this aberrant PI-kinase domain.

Here we describe a novel homology domain spanning ~500 amino acids N-terminal to the PI-kinase domain in the PIK-related and TRRAP subfamilies (Fig. 1b). The multiple alignment was constructed by iterative hidden Markov model searching using HMMER2 (see <http://hmmer.wustl.edu>) and manual alignment refinement. Because the middle portion in the multiple alignment is poorly conserved (see Fig. 1b), we were tempted to propose two domains. However, because all members contain conserved motifs over the entire region, we defined a single domain, called FAT, after representatives of the three main groups sharing the domain (FRAP, ATM, and TRRAP; see Fig. 1a). This domain is not found outside these subfamilies. Because the previously mentioned extreme C-terminal domain is also only found in these subfamilies, we call it FATC. The FAT and FATC domains only occur in combination, suggesting that they interact with each other. It is possible that they fold together in a configuration that ensures proper function of the PI-kinase domain, which is wedged in between the FAT and FATC domains. The FATC domain is probably too small (~35 amino acids) to fold independently, but because it is more conserved than the FAT domain (34% versus 16% average identity), it could be more important for catalytic activity than the FAT domain.

In the FRAP and TRRAP groups it is quite clear from the tree in Fig. 1a which *Caenorhabditis elegans* proteins (Q18667 and O01438) are the likely orthologs to human counterparts. For the ATM group, however, it is unclear whether the

C. elegans protein Q22258 is orthologous to ATM or ATR; perhaps it is orthologous to both.

The functions of the FAT and FATC domains still need to be elucidated experimentally. Data from deletion experiments regarding the functional importance of the N-terminal region in ATM (Refs 19,20) and RAD3 (Ref. 15) have proved contradictory. The region upstream of the PI-kinase domain, including the FAT domain, contains numerous regions of low sequence complexity. These are not of a standard type, such as coiled-coil, but many are enriched in leucine and glutamate. The FAT domain has diverged much faster than the catalytic PI-kinase domain (16% versus 28% average identity). Such sequence properties are typically found in proteins with an extended, non-globular structure, or proteins that form multimeric protein complexes²⁴. We therefore speculate that the FAT domain could be of importance as a structural scaffold or as a protein-binding domain, or both.

References

- 1 Majerus, P.W. *et al.* (1990) Recent insights in phosphatidylinositol signaling. *Cell* 63, 459–465
- 2 Carr, A.M. (1997) Control of cell cycle arrest by the Mec1sc/Rad3sp DNA structure checkpoint pathway. *Curr. Opin. Genet. Dev.* 7, 93–98
- 3 Hoekstra, M.F. (1997) Responses to DNA damage and regulation of cell cycle checkpoints by the ATM protein kinase family. *Curr. Opin. Genet. Dev.* 7, 170–175
- 4 Hartley, K.O. *et al.* (1995) DNA-dependent protein kinase catalytic subunit: a relative of phosphatidylinositol 3-kinase and the ataxia telangiectasia gene product. *Cell* 82, 849–856
- 5 Bentley, N.J. *et al.* (1996) The *Schizosaccharomyces pombe* rad3 checkpoint gene. *EMBO J.* 15, 6641–6651
- 6 Brunn, G.J. *et al.* (1996) Direct inhibition of the signaling functions of the mammalian target of rapamycin by the phosphoinositide 3-kinase inhibitors, wortmannin and LY294002. *EMBO J.* 15, 5256–5267
- 7 Rotman, G. and Shiloh, Y. (1998) ATM: from gene to function. *Hum. Mol. Genet.* 7, 1555–1563
- 8 Zakian, V.A. (1995) ATM-related genes: what do they tell us about functions of the human gene? *Cell* 82, 685–687
- 9 Ziv, Y. *et al.* (1997) Recombinant ATM protein complements the cellular A-T phenotype. *Oncogene* 15, 159–167
- 10 Keith, C.T. and Schreiber, S.L. (1995) PIK-related kinases: DNA repair, recombination, and cell cycle checkpoints. *Science* 270, 50–51
- 11 Kato, R. and Ogawa, H. (1994) An essential gene, ESR1, is required for mitotic cell growth, DNA repair and meiotic recombination in *Saccharomyces cerevisiae*. *Nucleic Acids Res.* 22, 3104–3112
- 12 Meyn, M.S. (1995) Ataxia-telangiectasia and cellular responses to DNA damage. *Cancer Res.* 55, 5991–6001
- 13 Savitsky, K. *et al.* (1995) A single ataxia telangiectasia gene with a product similar to PI-3 kinase. *Science* 268, 1749–1753
- 14 Carpenter, C.L. and Cantley, L.C. (1996) Phosphoinositide kinases. *Curr. Opin. Cell Biol.* 8, 153–158
- 15 Chapman, C.R. *et al.* (1999) Requirement of sequences outside the conserved kinase domain of fission yeast Rad3p for checkpoint control. *Mol. Biol. Cell* 10, 3223–3238
- 16 Bornberg-Bauer, E. *et al.* (1998) Computational approaches to identify leucine zippers. *Nucleic Acids Res.* 26, 2740–2746
- 17 Montecucco, A. *et al.* (1998) DNA ligase I is recruited to sites of DNA replication by an interaction with proliferating cell nuclear antigen: identification of a common targeting mechanism for the assembly of replication factories. *EMBO J.* 17, 3786–3795
- 18 McMahon, S.B. *et al.* The novel ATM-related protein TRRAP is an essential cofactor for the c-Myc and E2F oncoproteins. *Cell* (1998) 94, 363–374
- 19 Baskaran, R. *et al.* (1997) Ataxia telangiectasia mutant protein activates c-Abl tyrosine kinase in response to ionizing radiation. *Nature* 387, 516–519
- 20 Morgan, S.E. and Kastan, M.B. (1997) p53 and ATM: cell cycle, cell death, and cancer. *Adv. Cancer Res.* 71, 1–25
- 21 Bateman, A. *et al.* (1999) Pfam 3.1: 1313 multiple alignments and profile HMMs match the majority of proteins. *Nucleic Acids Res.* 27, 260–262
- 22 Galtier, N. *et al.* (1996) SEAVIEW and PHYLO-WIN: two graphic tools for sequence alignment and molecular phylogeny. *Comput. Appl. Biosci.* 12, 543–548
- 23 Thompson, J.D. *et al.* (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* 22, 4673–4680
- 24 Wright, P.E. and Dyson, H.J. (1999) Intrinsically unstructured proteins: re-assessing the protein structure–function paradigm. *J. Mol. Biol.* 293, 321–331

ROBERTA BOSOTTI AND ANTONELLA ISACCHI

Dept of Biology, Pharmacia & Upjohn, Viale Pasteur 10, 20014, Nerviano (MI), Italy.

ERIK L.L. SONNHAMMER

Center for Genomics Research, Karolinska Institutet, 171 77 Stockholm, Sweden.
Email: Erik.Sonnhammer@cgr.ki.se

Nuclear receptors arose from pre-existing protein modules during evolution

Nuclear receptors (NRs) are ligand-modulated transcription factors comprising multiple domains that include a highly conserved DNA-binding domain (DBD) and a ligand-binding domain (LBD). Here, we support the hypothesis that nuclear receptors arose in evolution from a fusion event linking pre-existing and independent protein modules^{1,2} (see also: <http://www.biomednet.com/hmsbeagle/03/cutedge/day1.htm>).

One view held for the origin of multidomain proteins is that they arose by DNA shuffling and rearrangement, bringing together pre-existing protein modules in new chimeric combinations to

create proteins with potentially new functions³. The origin of NRs might comply with this view. For example, the zinc-finger DNA-binding elements seen in the NR show a high degree of structural similarity to LIM (Lin-11, Isl-1 and Mec-3) and GATA (GATA-DNA-binding transcription factors) domains, which are present in both unicellular and multicellular organisms⁴. Therefore, it is believed that such zinc-finger DBDs might well have evolved from LIM and GATA proteins. However, a candidate LBD precursor has remained elusive. Our analyses provide evidence that Pex11p, a protein found in all major eukaryotic

Kingdoms, could be an ancient relative of the LBD.

Pex11p is a peroxisomal membrane protein of 235 amino acid residues implicated in peroxisome proliferation in the yeast *Saccharomyces cerevisiae*^{5,6} and in human cells^{7,8}. Although its precise function is unknown (for a critical discussion, see Ref. 9), Pex11p-deficient *S. cerevisiae* also displays a marked inability to degrade fatty acids (E.H. Hettema and C.W.T. van Roermund, unpublished). To obtain additional clues as to the possible function of Pex11p, we searched for similarities to other proteins. By combining both BLAST2 searches and CLUSTALX analyses, we observed a highly significant amino acid sequence similarity (30% identity, 50% similarity) between amino acids 2–187 of Pex11p and the LBD of NRs, in particular the peroxisome